

January 2011

# I/LA28-A2

Quality Assurance for Design Control and Implementation of Immunohistochemistry Assays; Approved Guideline—Second Edition

This document provides guidelines for the development of validated diagnostic, prognostic, and predictive immunohistochemical assays.

A guideline for global application developed through the Clinical and Laboratory Standards Institute consensus process.

## **Clinical and Laboratory Standards Institute**

Setting the standard for quality in clinical laboratory testing around the world.

The Clinical and Laboratory Standards Institute (CLSI) is a not-for-profit membership organization that brings together the varied perspectives and expertise of the worldwide laboratory community for the advancement of a common cause: to foster excellence in laboratory medicine by developing and implementing clinical laboratory standards and guidelines that help laboratories fulfill their responsibilities with efficiency, effectiveness, and global applicability.

### **Consensus Process**

Consensus—the substantial agreement by materially affected, competent, and interested parties—is core to the development of all CLSI documents. It does not always connote unanimous agreement, but does mean that the participants in the development of a consensus document have considered and resolved all relevant objections and accept the resulting agreement.

### **Commenting on Documents**

CLSI documents undergo periodic evaluation and modification to keep pace with advancements in technologies, procedures, methods, and protocols affecting the laboratory or health care.

CLSI's consensus process depends on experts who volunteer to serve as contributing authors and/or as participants in the reviewing and commenting process. At the end of each comment period, the committee that developed the document is obligated to review all comments, respond in writing to all substantive comments, and revise the draft document as appropriate.

Comments on published CLSI documents are equally essential, and may be submitted by anyone, at any time, on any document. All comments are addressed according to the consensus process by a committee of experts.

### **Appeals Process**

If it is believed that an objection has not been adequately addressed, the process for appeals is documented in the CLSI Standards Development Policies and Process document.

All comments and responses submitted on draft and published documents are retained on file at CLSI and are available upon request.

### Get Involved—Volunteer!

Do you use CLSI documents in your workplace? Do you see room for improvement? Would you like to get involved in the revision process? Or maybe you see a need to develop a new document for an emerging technology? CLSI wants to hear from you. We are always looking for volunteers. By donating your time and talents to improve the standards that affect your own work, you will play an active role in improving public health across the globe.

For further information on committee participation or to submit comments, contact CLSI.

Clinical and Laboratory Standards Institute 950 West Valley Road, Suite 2500 Wayne, PA 19087 USA P: 610.688.0100 F: 610.688.0700 www.clsi.org standard@clsi.org

	I/LA28-A2
	Vol. 31 No. 4
ISBN 1-56238-745-6	Replaces MM04-A
ISSN 0273-3099	Vol. 19 No. 26

### Quality Assurance for Design Control and Implementation of Immunohistochemistry Assays; Approved Guideline—Second Edition

Volume 31 Number 4

Stephen M. Hewitt, MD, PhD Max Robinowitz, MD Steven A. Bogen, MD, PhD Allen M. Gown, MD Krishan L. Kalra, PhD Christopher N. Otis, MD Betsy Spaulding Clive R. Taylor, MD, DPhil

### Abstract

Immunohistochemistry is an analytical technique that applies an antibody reagent to detect and visualize an antigen in cytological and surgical pathology microscopy specimens in the context of histomorphology and cytomorphology. The clinical-pathological interpretation of the presence and patterns of the antibody-antigen reactions is performed in a manner similar to other molecular pathology assays. Immunohistochemistry is used in diagnostic pathology for diagnosis, determination of prognosis, and predictive assays for response to therapy. Accurate and reproducible results require quality assurance of the total test system including the design control of the reagents and the preexamination (preanalytical), examination (analytical), and postexamination (postanalytical) interpretation steps (processes) of the assay to ensure its clinical applicability. This guideline focuses on validation of immunohistochemistry assays on formalin-fixed, paraffin-embedded pathology material. The audience for this guideline includes the assay developer, the reagent supplier, laboratory histotechnologist who performs the assay, and the laboratory director/pathologist who implements and interprets the assay.

Clinical and Laboratory Standards Institute (CLSI). *Quality Assurance for Design Control and Implementation of Immunohistochemistry Assays; Approved Guideline—Second Edition.* CLSI document I/LA28-A2 (ISBN 1-56238-745-6). Clinical and Laboratory Standards Institute, 950 West Valley Road, Suite 2500, Wayne, Pennsylvania 19087 USA, 2011.

The Clinical and Laboratory Standards Institute consensus process, which is the mechanism for moving a document through two or more levels of review by the health care community, is an ongoing process. Users should expect revised editions of any given document. Because rapid changes in technology may affect the procedures, methods, and protocols in a standard or guideline, users should replace outdated editions with the current editions of CLSI documents. Current editions are listed in the CLSI catalog and posted on our website at www.clsi.org. If your organization is not a member and would like to become one, and to request a copy of the catalog, contact us at: Telephone: 610.688.0100; Fax: 610.688.0700; E-Mail: customerservice@clsi.org; Website: www.clsi.org.



Number 4

I/LA28-A2

Copyright <sup>©</sup>2011 Clinical and Laboratory Standards Institute. Except as stated below, any reproduction of content from a CLSI copyrighted standard, guideline, companion product, or other material requires express written consent from CLSI. All rights reserved. Interested parties may send permission requests to permissions@clsi.org.

CLSI hereby grants permission to each individual member or purchaser to make a single reproduction of this publication for use in its laboratory procedure manual at a single site. To request permission to use this publication in any other manner, e-mail permissions@clsi.org.

### **Suggested Citation**

CLSI. Quality Assurance for Design Control and Implementation of Immunohistochemistry Assays; Approved Guideline—Second Edition. CLSI document I/LA28-A2. Wayne, PA: Clinical and Laboratory Standards Institute; 2011.

**Proposed Guideline** 

July 1997

**Approved Guideline** December 1999

Approved Guideline—Second Edition January 2011

ISBN 1-56238-745-6 ISSN 0273-3099

Volume 31

### **Committee Membership**

### Area Committee on Immunology and Ligand Assay

Ronald J. Whitley, PhD Chairholder University of Kentucky Medical Center Lexington, Kentucky, USA

Robert F. Vogt, Jr., PhD Vice-Chairholder **Centers for Disease Control and** Prevention Atlanta, Georgia, USA

Dorothy J. Ball, PhD Abbott Irving, Texas, USA

Bernard C. Cook, PhD, DABCC, FACB Beckman Coulter, Inc. Chaska, Minnesota, USA

Joan H. Howanitz, MD SUNY Brooklyn Brooklyn, New York, USA

### Subcommittee on Quality Assurance for Immunohistochemistry

Stephen M. Hewitt, MD, PhD Co-Chairholder National Institutes of Health. National Cancer Institute Bethesda, Maryland, USA

### Max Robinowitz, MD **Co-Chairholder** FDA Ctr. for Devices/Rad. Health Rockville, Maryland, USA

Steven A. Bogen, MD, PhD Boston University School of Medicine Boston, Massachusetts, USA

Allen M. Gown, MD PhenoPath Laboratories Seattle, Washington, USA

Krishan L. Kalra, PhD **BioGenex** Laboratories San Ramon, California, USA Marilyn M. Lightfoote, MD, PhD FDA Center for Devices and Radiological Health Silver Spring, Maryland, USA

Tom H. Stahlberg PerkinElmer Finland Oy Turku, Finland

Robert W. Veltri, PhD Johns Hopkins Hospital Baltimore, Maryland, USA

### Advisors

W. Harry Hannon, PhD Centers for Disease Control and Prevention Atlanta, Georgia, USA

Robin G. Lorenz, MD, PhD University of Alabama at Birmingham Birmingham, Alabama, USA

Christopher N. Otis, MD Baystate Medical Center Springfield, Massachusetts, USA

**Betsy Spaulding** Dako North America Carpinteria, California, USA

Clive R. Taylor, MD, DPhil USC Keck School of Medicine Los Angeles, California, USA

### Advisors

Odile David, MD **Tulane University** New Orleans, Louisiana, USA

Bharat Jasani, MD Cardiff University Cardiff, United Kingdom

Hans Lyon, MD Hvidovre Hospital Hvidovre, Denmark

Timothy J. O'Leary, MD, PhD Department of Veterans Affairs Washington, DC, USA

Gerald E. Marti, MD, PhD FDA Center for Biologics Evaluation and Research Bethesda, Maryland, USA

Per N. J. Matsson, PhD Phadia AB Uppsala, Sweden

Robert M. Nakamura, MD Scripps Clinic & Research Foundation La Jolla, California, USA

Jelili Ojodu, MPH Association of Public Health Laboratories Silver Spring, Maryland, USA

Philip R. Wyatt, MD, PhD York Central Hospital, Toronto North York, Ontario, Canada

Elizabeth Sheppard, HT(ASCP) Ventana Medical Systems Inc. Tucson, Arizona, USA

Jacqueline B. Welch, MHS, MT(ASCP)SBB FDA/CDRH/OC/DOEA Rockville, Maryland, USA

### Staff

Clinical and Laboratory Standards Institute Wayne, Pennsylvania, USA

Lois M. Schmidt, DA Vice President, Standards Development

Ron S. Quicho Staff Liaison

Melissa A. Lewis, ELS Editorial Manager

Megan P. Larrisey, MA Assistant Editor

Number 4

I/LA28-A2

### Acknowledgment

Clinical and Laboratory Standards Institute (CLSI) gratefully acknowledges the contribution of the following experts in developing Appendixes A1 and A2 of this revised guideline:

Andre J. Balaton, MD Centre de Pathologie Bievres, France

Patrick Fitzgibbons, MD College of American Pathologists Northfield, Illinois, USA

Shanti Gomatam, PhD US Food and Drug Administration Silver Spring, Maryland, USA

Eunyoung Kim, MS US Food and Drug Administration Silver Spring, Maryland, USA

Marina Kondratovich, PhD US Food and Drug Administration Silver Spring, Maryland, USA

Samir Lababidi, PhD US Food and Drug Administration Rockville, Maryland, USA

Kristen Meier, PhD US Food and Drug Administration Silver Spring, Maryland, USA

Keith Miller UK NEQAS – ICC & FISH, UCL – Advanced Diagnostics London, United Kingdom

Robert Y. Osamura, MD Tokai University School of Medicine Kanagawa, Japan

Gene Pennello, PhD US Food and Drug Administration Silver Spring, Maryland, USA

Estelle Russek-Cohen, PhD US Food and Drug Administration Silver Spring, Maryland, USA

Betsy Spaulding Dako North America Carpenteria, California, USA

Robert Veltri, PhD Johns Hopkins University Hospital Baltimore, Maryland, USA

Volume 31	I/LA28-A2

### Contents

Abstra	ct		i
Comm	ittee Me	mbership	iii
Forewo	ord		ix
1	Scope.		1
2	Introdu	iction	1
3	Standard Precautions		
4	Terminology		3
		A Note on Terminology Definitions Nomenclature Abbreviations and Acronyms entific Theory and Principles of the Design and Development of nemistry Assays	3 13 13
5		Control Process for Immunohistochemistry Assay Discovery, Development, nical Implementation	15
	<ul> <li>5.1</li> <li>5.2</li> <li>5.3</li> <li>5.4</li> <li>5.5</li> <li>5.6</li> <li>5.7</li> <li>5.8</li> </ul>	Verification and Validation of the Design Control Process of an Immunohistochemistry Assay or Component Intended Use and Indications for Use of an Immunohistochemistry Assay Discovery and Development of the Antigen and Antibody Reagents of an Immunohistochemistry Assay Total Test System Common to All Finished Immunohistochemical Assays Verification During Each Step of the Design Control Process Validation of the Total Test System Finished Assay by the Immunohistochemistry Test Developer to Confirm That User Requirements Were Met Lot-to-Lot (Batch-to-Batch) Acceptance Process by Test Developer and End User Commercialized Immunohistochemistry Assays	17 18 18 18 18 18
	5.8 5.9 5.10 5.11	Quality Assurance and Quality Control Clinical Effectiveness of Device Device Failure Reporting to Stakeholders and Regulatory Bodies	20 20
6	Specim	en Requirements for Immunohistochemistry Assays	22
	<ul> <li>6.1</li> <li>6.2</li> <li>6.3</li> <li>6.4</li> <li>6.5</li> <li>6.6</li> <li>6.7</li> </ul>	Specimen Acquisition Specimen Handling Fixation Postfixation Processing Processing Tissue Into Paraffin Cytological Specimens Tissue Processing Problem Troubleshooting	23 25 31 31 36
7		Properties of the Terret Antigen	
	7.1	Properties of the Target Antigen	

Number 4

### **Contents (Continued)**

8	Primar	Primary Antibody	
	8.1	Immunogen (Epitope)	40
	8.2	Antibodies	40
	8.3	Characterization of Antibodies	
	8.4	Tissue Microarrays	46
9	9 Detection Methodologies and Chemistries		
	9.1	Direct Method—One Antibody Step (see Figure 3A)	50
	9.2	Indirect Method (see Figure 3B)	
	9.3	Other Types of Indirect Methods	51
	9.4	Multiple Staining Methods—Staining More Than One Target Antigen per	<i></i>
	0.5	Specimen	
	9.5 9.6	Quick Staining Methods           Frozen Tissue Specimen Immunofluorescent Staining Methods	
10	Desigr	Principles for Processing and Immunostaining Steps	55
	10.1	Pretreatment—Deparaffinization, Rehydration, and Antigen Retrieval	
	10.2	Blocking	
	10.3	Application of Primary Antibody	
	10.4 10.5	Application of Secondary Antibody and Detection System Completion of Assay	
	10.5	Completion of Assay	
	10.0	Interpretation of Staining	
	10.7	Automation of Staining	
11	Desigr	Principles for Interpretation of Immunohistochemistry Assays	
	11.1	Interpretation of Immunostaining	
	11.1	Expected vs Observed Results	
	11.2	Staining Patterns	
	11.4	Expected vs Observed Results	
	11.5	Methods for Determination of Appropriate Controls	
	11.6	Validation—Establishing Diagnostic Performance	
	11.7	Controls	
	11.8	Observation of the Result	76
	11.9	Scoring Systems (See Appendix A1, Statistical Points to Consider When	
		Evaluating the Precision of Immunohistochemistry Assays)	//
Part 2	. The Im	plementation of Immunohistochemistry Assays by Pathologists and Technologists	81
12	Assay	Verification and Validation in the End-User Laboratory	81
	12.1	Directed Guidance for End-User Laboratories on Verification	
		(See Appendix A1, Statistical Points to Consider When Evaluating the Precision	
		of Immunochemistry Assays)	
	12.2	Directed Guidance for End-User Laboratory on Validation	
	12.3	Surveillance and Distribution Analysis of Laboratory Results	
	12.4	Quality Assurance	
13	Selecti	on of Immunohistochemistry Assay by Surgical Pathologist	85
	13.1	Goal for Any New Clinical Test	
	13.2	Immunohistochemistry Validation	
	13.3	Cost-Benefit of an Immunohistochemistry Test	86

Vol	ume	31

**Contents (Continued)** 

	13.4	Selection of Antibody Choices and Antibody Panels by Pathologists	86
14	Repor	ting of Results by Pathologists	87
	14.1	Specific Suggestions and Concerns About Reporting of Results	88
15	Reage	nt Handling by End-User Laboratory	89
Part 3.	Regula	tion and Future Developments in Immunohistochemistry Assays	90
16	Regula	atory Issues	90
	16.1 16.2 16.3 16.4	International Regulations National Regulations Local Regulations Professional Association Practice Guidelines	90 91
17	Allied	Assays	91
	17.1	Independent Validation of Immunohistochemistry by Reflex Fluorescence <i>In Situ</i> Hybridization and Other Tests	91
18	The F	uture of Immunohistochemistry	92
	18.1 18.2	Expected Changes Anticipated Changes in the Procurement, Handling, and Processing of Surgical	
	18.3	Tissue Predictive Biomarker Discovery and Development	
Refere	nces	· · · ·	
Appen	dix A1.	Statistical Points to Consider When Evaluating the Precision of Immunohistochemis	stry
Appen	dix A2.	Data Recording Sheets, Intermediate Precision Experiment, Continuous Assay	118
		Comparison of the Characteristics of Immunoassays Such as Enzyme-Linked nt Assay and Immunohistochemistry	125
		The Essential Components of the Total Test System for Immunohistochemistry	129
		The US Food and Drug Administration's Risk-Based Regulatory Evaluation of hemistry Applications	133
The Q	uality M	lanagement System Approach	136
Relate	d CLSI	Reference Materials	138

Number 4

Volume 31

I/LA28-A2

### Foreword

This document is a revision of the previous CLSI document MM04. The goal for the revision is to maintain the consensus recommendations of MM04 while incorporating new information that has accrued since the 1999 publication. The emphasis of this guideline is the application of immunohistochemistry (IHC) and immunocytochemistry (ICC) for the study of human tumor specimens. Other uses, such as identification of microorganisms in specimens, are not addressed.

IHC is just one of the methods intended to assess the diagnosis, prognosis, and prediction of therapy for human tumor specimens. However, IHC and ICC have unique utility through their power to apply immunological methods to localize macromolecular targets within the context of standard histomorphology and cytological morphology. This is particularly useful in tumors that are morphologically anaplastic or are heterogeneous in their cellularity and differentiation.

In the era of personalized or individualized medicine, physicians hope to identify the therapies that are most likely to be effective and avoid therapies that are likely to have unwanted side effects. In order to achieve personalized medicine (ie, individualized medicine), there is a global effort by pharmaceutical and medical device researchers and developers to apply fit-for-purpose method development in the successful identification of biomarkers. There is also an effort to ensure these biomarkers are accurate and reliable for the calculation of the prognosis of the tumor of individual patients, and for predicting optimal response to individualized therapies.<sup>1</sup>

I/LA28-A2 advances the MM04 recommendations for performance of immunohistochemical assays to promote a better understanding of the requirements, capabilities, and limitations of these diagnostic methods; to improve their intra- and interlaboratory reproducibility; and to improve their positive and negative predictive values in diagnosis of disease.

MM04 was revised as I/LA28-A2 because ICC and IHC are based on immunological detection methods, whereas CLSI molecular methods documents are concerned with nucleic acid-based assays. In addition, I/LA28-A2 is intended as a companion document to CLSI documents I/LA23<sup>2</sup> and I/LA21.<sup>3</sup>

This document contains detailed recommendations about preexamination (preanalytical) specimen handling, processing, and preparation. Tissue specimens, especially formalin-fixed, paraffin-embedded tissue are widely used in a variety of different assays today. The inclusion of these preexamination (preanalytical) factors in this document is based on previously published literature and consensus best practices.

Additionally, this document contains recommendations on design and statistical analysis of experiments for estimating the precision of IHC assay results.

### **Key Words**

Anatomical pathology, antigen retrieval, biorepository, formalin fixation, immunocytochemistry, immunohistochemistry, surgical pathology, validation, verification

Number 4

Volume 31

I/LA28-A2

### Quality Assurance for Design Control and Implementation of Immunohistochemistry Assays; Approved Guideline—Second Edition

### 1 Scope

The purpose of this document is to provide guidance to all of the stakeholders involved with design and implementation of immunohistochemistry (IHC) assays. It is intended for use by all clinical and reference laboratories performing immunocytochemical assays on cytological preparations or immunohistochemical assays on surgical pathology specimens, for the manufacturers of commercial reagents and test kits, and for individuals and organizations involved in the development and implementation of laboratory quality assurance (QA) programs for these assays.

This guideline presents information on the total product life cycle of the discovery, design, development, verification, and analytical and clinical validation of IHC and immunocytochemistry (ICC) reagents, kits, and systems. Its emphasis is that accurate and reliable IHC and ICC results require attention to the total test system of the assay.

### 2 Introduction

In preparing this guideline, the subcommittee considered the following needs of stakeholders in the total life cycle of IHC tests:

- Discovery of antigens and antibodies of biological and clinical interest
- Development of research and investigational IHC assays
- Translation to clinical applications: the intended use and indications for use of the test
- Design, verification, and validation of an assay for clinical use
- QA recommendations for laboratory-developed assays and commercialized assays by manufacturers, and for routine clinical laboratory users of IHC assays
- Proficiency testing and other user issues
- Regulation, harmonization, and standardization

The document emphasizes how to address the unique challenges to optimize immunological methodologies applied to histological preparations and cytology.

This document is divided into three parts: Part 1 is the scientific theory and principles of the design and development of IHC assays; Part 2 is the implementation of IHC tests by pathologists and technologists; and Part 3 is the regulation issues and future developments in IHC assays. Figure 1 shows the participants throughout the total IHC life cycle.