

First edition
2017-03

Microbiology of the food chain — Horizontal method for determination of hepatitis A virus and norovirus using real-time RT-PCR —

Part 1: Method for quantification

*Microbiologie dans la chaîne alimentaire — Méthode horizontale
pour la recherche des virus de l'hépatite A et norovirus par la
technique RT-PCR en temps réel —*

Partie 1: Méthode de quantification



Reference number
ISO 15216-1:2017(E)

© ISO 2017

This is a preview of "ISO 15216-1:2017". [Click here to purchase the full version from the ANSI store.](#)



COPYRIGHT PROTECTED DOCUMENT

© ISO 2017, Published in Switzerland

All rights reserved. Unless otherwise specified, no part of this publication may be reproduced or utilized otherwise in any form or by any means, electronic or mechanical, including photocopying, or posting on the internet or an intranet, without prior written permission. Permission can be requested from either ISO at the address below or ISO's member body in the country of the requester.

ISO copyright office
Ch. de Blandonnet 8 • CP 401
CH-1214 Vernier, Geneva, Switzerland
Tel. +41 22 749 01 11
Fax +41 22 749 09 47
copyright@iso.org
www.iso.org

This is a preview of "ISO 15216-1:2017". Click here to purchase the full version from the ANSI store.

Contents

	Page
Foreword	v
Introduction	vi
1 Scope	1
2 Normative references	1
3 Terms and definitions	1
4 Principle	3
4.1 Virus extraction.....	3
4.2 RNA extraction.....	4
4.3 Real-time RT-PCR.....	4
4.4 Control materials.....	4
4.4.1 Process control virus.....	4
4.4.2 Double-stranded DNA (dsDNA) control.....	4
4.4.3 EC RNA control.....	4
4.5 Test results.....	5
5 Reagents	5
5.1 General.....	5
5.2 Reagents used as supplied.....	5
5.3 Prepared reagents.....	6
6 Equipment and consumables	7
7 Sampling	9
8 Procedure	9
8.1 General laboratory requirements.....	9
8.2 Virus extraction.....	9
8.2.1 Process control virus material.....	9
8.2.2 Negative process control.....	9
8.2.3 Food surfaces.....	9
8.2.4 Soft fruit, leaf, stem and bulb vegetables.....	9
8.2.5 Bottled water.....	10
8.2.6 Bivalve molluscan shellfish.....	11
8.3 RNA extraction.....	11
8.4 Real-time RT-PCR.....	12
8.4.1 General requirements.....	12
8.4.2 Real-time RT-PCR analysis.....	12
9 Interpretation of results	14
9.1 General.....	14
9.2 Construction of standard curves.....	14
9.3 Calculation of RT-PCR inhibition.....	15
9.4 Calculation of extraction efficiency.....	15
9.5 Sample quantification.....	16
10 Expression of results	17
11 Precision	17
11.1 Interlaboratory study.....	17
11.2 Repeatability.....	17
11.3 Reproducibility limit.....	18
12 Test report	18
Annex A (normative) Diagram of procedure	19
Annex B (normative) Composition and preparation of reagents and buffers	20
Annex C (informative) Real-time RT-PCR mastermixes and cycling parameters	23

This is a preview of "ISO 15216-1:2017". [Click here to purchase the full version from the ANSI store.](#)

Annex D (informative) Real-time RT-PCR primers and hydrolysis probes for the detection of HAV, norovirus GI and GII and mengo virus (process control)	24
Annex E (informative) Growth of mengo virus strain MC₀ for use as a process control	27
Annex F (informative) RNA extraction using the NucliSENS® system	28
Annex G (informative) Generation of dsDNA control stocks	30
Annex H (informative) Generation of EC RNA stocks	33
Annex I (informative) Typical optical plate layout	35
Annex J (informative) Method validation studies and performance characteristics	37
Bibliography	48

This is a preview of "ISO 15216-1:2017". [Click here to purchase the full version from the ANSI store.](#)

Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights. Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see www.iso.org/patents).

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation on the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT) see the following URL: www.iso.org/iso/foreword.html.

This document was prepared by the European Committee for Standardization (CEN) Technical Committee CEN/TC 275, in collaboration with ISO Technical Committee ISO/TC 34, *Food products*, Subcommittee SC 9, *Microbiology*, in accordance with the agreement on technical cooperation between ISO and CEN (Vienna Agreement).

This first edition cancels and replaces ISO/TS 15216-1:2013, which has been technically revised with the following changes:

- use of linear dsDNA molecules for quantification prescribed;
- use of a suitable buffer for dilution of control materials prescribed;
- change to the method for generating process control virus RNA for the standard curve;
- addition of breakpoints with defined temperature and time parameters in the extraction methods;
- change in terminology from amplification efficiency to RT-PCR inhibition;
- addition of extra real-time RT-PCR reactions for negative controls;
- addition of precision data and results of interlaboratory study.

A list of all parts in the ISO 15216 series can be found on the ISO website.

Introduction

Hepatitis A virus (HAV) and norovirus are important agents of food-borne human viral illness. No routine methods exist for culture of norovirus, and HAV culture methods are not appropriate for routine application to food matrices. Detection is therefore reliant on molecular methods using the reverse-transcriptase polymerase chain reaction (RT-PCR). As many food matrices contain substances that are inhibitory to RT-PCR, it is necessary to use an extraction method that produces highly clean RNA preparations that are fit for purpose. For food surfaces, viruses are removed by swabbing. For soft fruit, leaf, stem and bulb vegetables, virus extraction is by elution with agitation followed by precipitation with PEG/NaCl. For bottled water, adsorption and elution using positively charged membranes followed by concentration by ultrafiltration is used and for bivalve molluscan shellfish (BMS), viruses are extracted from the tissues of the digestive glands using treatment with a proteinase K solution. For all matrices that are not covered by this document, it is necessary to validate this method. All matrices share a common RNA extraction method based on virus capsid disruption with chaotropic reagents followed by adsorption of RNA to silica particles. Real-time RT-PCR monitors amplification throughout the real-time RT-PCR cycle by measuring the excitation of fluorescently labelled molecules. In real-time RT-PCR with hydrolysis probes, the fluorescent label is attached to a sequence-specific nucleotide probe that also enables simultaneous confirmation of target template. These modifications increase the sensitivity and specificity of the real-time RT-PCR method, and obviate the need for additional amplification product confirmation steps post real-time RT-PCR. Due to the complexity of the method, it is necessary to include a comprehensive suite of controls. The method described in this document enables quantification of levels of virus RNA in the test sample. A schematic diagram of the testing procedure is shown in [Annex A](#).

The main changes, listed in the Foreword, introduced in this document compared to ISO/TS 15216-1:2013 are considered as minor (see ISO 17468).